

This is a repository copy of *Multiple introductions and environmental factors affecting the establishment of invasive species on a volcanic island*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/84521/>

Version: Published Version

Article:

Novo, M., Cunha, L., Maceda-Veiga, A. et al. (6 more authors) (2015) Multiple introductions and environmental factors affecting the establishment of invasive species on a volcanic island. *Soil Biology and Biochemistry*. pp. 89-100. ISSN 0038-0717

<https://doi.org/10.1016/j.soilbio.2015.02.031>

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



Multiple introductions and environmental factors affecting the establishment of invasive species on a volcanic island

M. Novo ^{a,*}, L. Cunha ^a, A. Maceda-Veiga ^a, J.A. Talavera ^b, M.E. Hodson ^c, D. Spurgeon ^d, M.W. Bruford ^a, A.J. Morgan ^a, P. Kille ^a

^a School of Biosciences, Cardiff University, UK

^b Departamento de Zoología, Universidad de la Laguna, Spain

^c Environment Department, University of York, UK

^d Centre for Ecology & Hydrology, Wallingford, UK

ARTICLE INFO

Article history:

Received 9 January 2015

Received in revised form

20 February 2015

Accepted 26 February 2015

Available online 11 March 2015

Keywords:

Earthworms

Invasive species

Azores

Altitude

Metals

Human introduction

ABSTRACT

Invasive species pose significant challenges to local biodiversity and ecosystem function, especially on islands. Understanding the factors affecting the establishment of invasive species and how these relate to their genetic background is crucial to improve our ability to manage biological invasions. Here, we performed a phylogeographic study of two cosmopolitan megascolecid earthworms of Asian origin: *Amyntas gracilis* and *Amyntas corticis* at 38 localities on São Miguel Island in the Azores archipelago (Portugal). Samples from putative source populations in China, Taiwan, Malaysia, as well as 'outlier' populations in USA, Mexico, Brazil and Spain were also included, resulting in a total of 565 earthworms genotyped at the mitochondrial cytochrome oxidase I (COI) and 16S ribosomal RNA genes. Soils were characterised for elemental composition, water holding capacity, organic matter content, texture and pH, and some habitat features were recorded. Both species showed a wide distribution across São Miguel and their abundances were negatively associated, suggesting spatial segregation/competition, with the parthenogenetic *A. corticis* being relatively more successful. The presence of multiple mitochondrial lineages within each species, one of them found exclusively in the Azores, suggests a complex invasion history. Environmental factors affected the establishment of the different lineages, with metal concentrations, topographical elevation and the degree of human influence being differently linked to their abundances. Lineage diversity was negatively correlated with metal concentrations. These results emphasise the importance of genetically characterising invasive species to better understand their invasion patterns.

© 2015 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Invasive species can adversely affect local biodiversity due to alterations of recipient ecosystems, impacts on native species, such as competition, predation or hybridisation, or as carriers of disease (Pejchar and Mooney, 2009). Global environmental change and globalisation of trade networks mean that introductions are more likely to occur and their proper prediction, prevention and management is crucial. Therefore understanding the influence of the environment, geography and genetic features on colonisation and

invasiveness of species should be a priority. Islands are particularly vulnerable to invasions, which are the main cause of population declines and species extinctions as well as having substantial socioeconomic impacts within these areas (Reaser et al., 2007). Invasive species have shown rapid adaptations to new biotic and abiotic environments and much of this evidence has been observed on islands, highlighting them as evolutionary hotspots (Mooney and Cleland, 2001).

Terrestrial invertebrates have been identified as one of the groups with the most profound ecological and economic impact through their roles in environmental processes that give rise to ecosystem services (Vila et al., 2010). However, attention has been focused on invasions above ground while soil organisms have been largely unexplored, even though their invasions may profoundly affect soil ecosystems (Hendrix et al., 2008). Alterations of the soil

* Corresponding author. School of Biosciences, Cardiff University, Museum Avenue, Main Building, CF310AT, Cardiff, UK. Tel.: +44 (0)29 208 74507.

E-mail address: novom@cf.ac.uk (M. Novo).

structure by earthworms can cause a cascade of ecological effects (Frelich et al., 2006). In the last decade several authors have studied the effects of introduced earthworms on temperate and tropical regions, showing their capacity to greatly modify nutrient and organic matter dynamics, above- and below-ground community structures, and soil structural properties; the role of humans in earthworm dispersal has also been shown to be significant (Hendrix et al., 2008 and references therein). A well-documented case concerns the invasion of European earthworms (Lumbricidae) in North America, which has resulted in a change in forest floor litter dynamics with resulting effects on ecosystem processes (e.g., Bohlen et al., 2004; Hale et al., 2005; Frelich et al., 2006). Less understood are the effects of megascolecid species, although Burtelow et al. (1998) showed that *Amyntas gracilis* in particular has impacted C and N fluxes in soils of the northeastern United States.

The Azores archipelago is located in the North Atlantic Ocean and comprises nine islands. Of these, São Miguel is the largest and is made up of active volcanic areas including fumarolic fields and cold and thermal springs with soil-diffuse degassing (Viveiros et al., 2009). Humans first populated the island in the late 1430's (Santos et al., 2005) and dramatic changes in land-use started after colonization. Native vegetation at low and middle altitudes became extinct or was modified and exotic plants and animals were introduced (Borges et al., 2006 and references therein). Among the Macaronesian archipelagos, the Azores contain fewer single-island endemic species (Amorim et al., 2012), which has been referred to as the 'Azores diversity enigma' (Carine and Schaefer, 2010). This relatively depauperate native species diversity could make the Azores archipelago more susceptible to successful invasions because more niches are unoccupied and, for example, 70% of the current flora and 58% of the arthropod fauna are exotic (Borges et al., 2006).

Most phylogeographic studies of earthworms to date have been carried out within the natural ranges of the species, including megascolecids (e.g., Chang and Chen, 2005; Chang et al., 2008; Buckley et al., 2011; Fernandez et al., 2011; Novo et al., 2011). Cameron et al. (2008) and Cunha et al. (2014) investigated the population genetic structure of invasive earthworms in the US and the Azores respectively, in relation to human-mediated dispersal and landscape features. Porco et al. (2013) and Shekhovtsov et al. (2014) have also applied DNA barcoding to study invasions of earthworms in the US and Kamchatka. More studies are needed to better understand the dynamic forces affecting earthworm invasions given the potential consequences for native species and ecosystems.

The Megascolecidae is the largest earthworm family and has a suggested Pangean origin (Jamieson et al., 2002). The genus *Amyntas* is thought to be native to the eastern Palearctic, with species being described as cosmopolitan, peregrine and invasive. *Amyntas corticis* and *A. gracilis* are amongst the most invasive earthworms on earth, mainly due to their inherent environmental plasticity, rapid growth, high reproductive performance and relatively large adult body size (Burtelow et al., 1998; Garcia and Fragoso, 2002). Additionally, some lineages of *A. corticis* including those from the Azores (unpublished observations by the authors) are parthenogenetic. This mode of reproduction can facilitate invasiveness (Terhivuo and Saura, 2006). For example, parthenogenetic species can found stable populations initiated by a single individual in novel environments. The benefit of uniparental reproduction is highest after long-distance dispersal (Baker, 1965, 1967), which would be the case following human introductions.

It is not presently known whether the introduction of megascolecid worms to the Azores was a single or multiple event(s); neither is it known whether the invaders originated from one or

more populations in the Eastern Palearctic (Hendrix et al., 2008). Given that the establishment on São Miguel Island by both *A. corticis* and *A. gracilis* is probably a relatively recent human-mediated event, they constitute an ideal model to test how the genetic background of an invader responds to local geography and the chemical characteristics of a new environment, in this case an island characterised by a volcanic-driven topography and geology. Physical features are known to affect earthworm distributions even on a very small scale (Rossi et al., 1997; Nuutinen et al., 1998; Hernandez et al., 2007; Jiménez et al., 2014).

Our aim was to study the phylogeography of megascolecids in São Miguel Island in the Azores (Portugal), placing their human introduction into a global context and exploring the likely factors affecting their establishment on this island. Specifically we aimed to: i) compare the haplotypes of Azorean *A. gracilis* and *A. corticis* to those of both species from other global sources (including Asian) in an attempt to detect different introduction events; ii) study the extent of genetic variability at the landscape scale in São Miguel by identifying genetic lineages within both species and their spatial distributions; iii) explore the relationships between the abundances of different lineages and environmental characteristics. Addressing these issues has allowed us to ascertain whether the invasion success of *Amyntas* on São Miguel is attributed to just one highly adapted genetic lineage per species, or to multiple lineages with potentially different adaptive capacities to spatially heterogeneous environmental conditions.

2. Materials and methods

2.1. Sampling

Samples were collected from throughout São Miguel Island (Azores, Portugal). Its area is 757 km² and includes three active volcanoes: Furnas (eastern end, ca. 1 million years old), Sete Cidades (western end, 550–750 thousand years old) and Fogo (center, ca. 350 thousand years old) (Gomes et al., 2006; Cunha et al., 2014). The oldest area of the island, to the east is ca. 4 million years old (Harris et al., 2013). The climate in São Miguel is oceanic and temperate being strongly influenced by altitude (Ricardo et al., 1977). The summits are higher in the eastern part (Pico da Vara, 1080 m) than in the western part (Eguas, 873 m), and the central part is the lowest (maximum of 400 m) (Louvat and Allègre, 1998). Mean annual temperatures range between 11.5 °C at the summit of Agua de Pau volcano (800–900 m) and 17.3 °C in Ponta Delgada (sea level). Annual rainfall and average relative humidity increase with elevation: rainfall ranges from 1000–1500 mm/yr in the driest part of the island (the south coast) to 3000 mm/yr at Lagoa do Fogo; relative humidity ranges from 77–78% on the coast to 87–88% at the highest elevation (Malucelli et al., 2006). Around 500 megascolecid earthworms were collected from 37 localities (Supplementary Table 1, Fig. 1B, C). The sampling effort was standardized in all the locations in order to record relative (but not absolute) abundances and to compare these between sites. These relative numbers are named abundances hereafter within the text. We sampled at least 8 quadrats (50 cm × 50 cm) randomly distributed in the sampling area that were separated by at least 3 m. Sampling effort was limited by time and people making the combination uniform among sites (generally 4 people digging for 60 min) with the search targeted to *Amyntas* specimens. All individuals were hand collected, washed in distilled water, weighted *in vivo* and subsequently fixed and preserved in ca. 96% EtOH. A portion of the integument (ca. 25 mg) was cleansed and preserved at –20 °C for DNA extraction. Samples from other countries in the species' global distribution, including from the presumed native range (Malaysia: MAL, China: YN, HB; Taiwan: TW), were donated

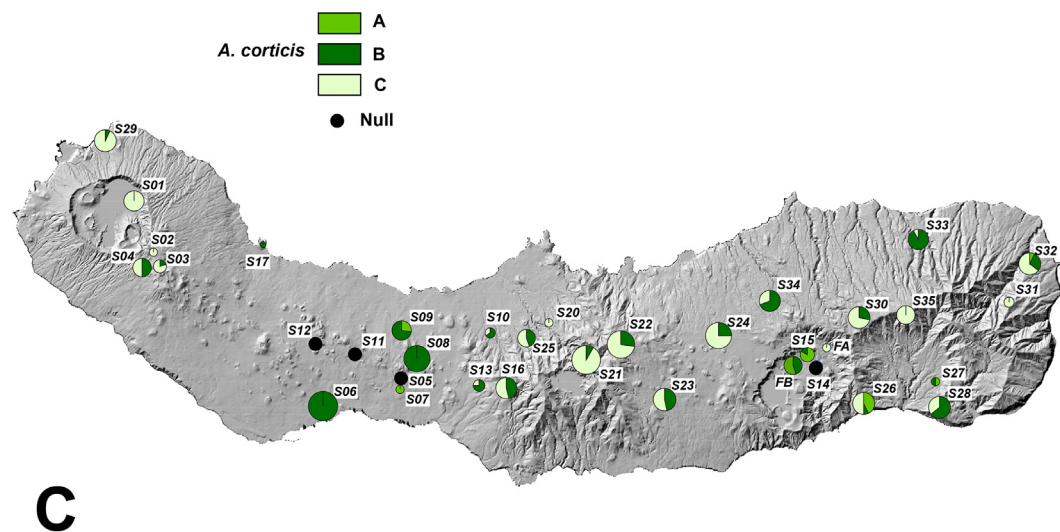
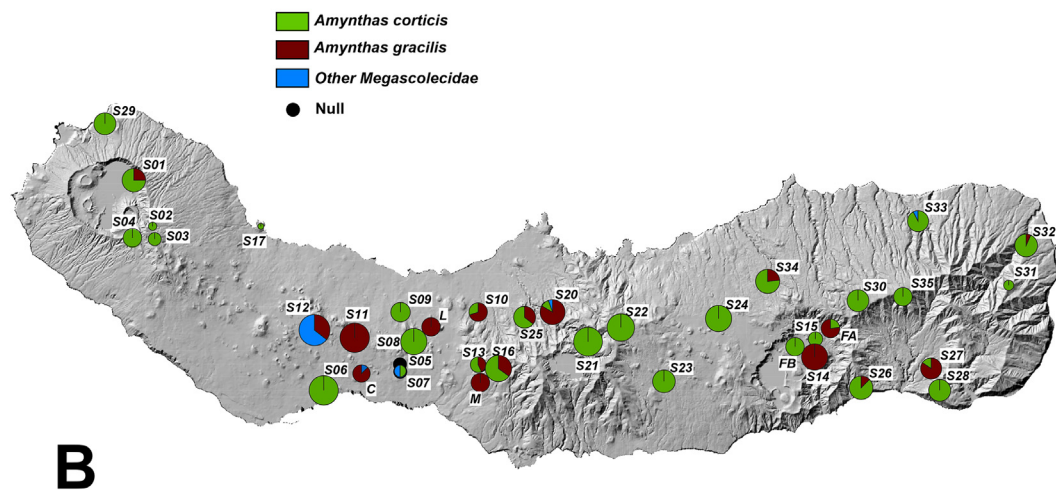
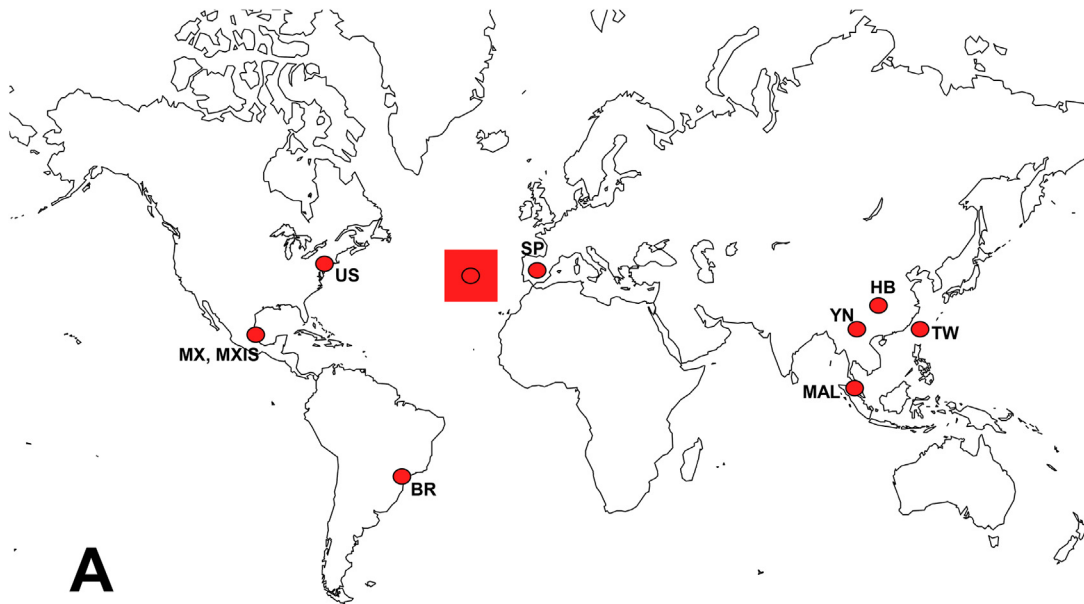


Fig. 1. Geographic distribution of the samples analysed. A: Sample sites outside Azores: São Miguel location is represented by a square. B: Sample sites within São Miguel Island (Azores). Proportion of different species found is shown. C: Distribution of haplotypes of the COI gene found within *Amynthus corticis*.

by colleagues (see [Supplementary Table 2](#), [Fig 1A](#) for sampling locations).

2.2. DNA extraction, gene amplification and sequencing

Total genomic DNA was extracted using the DNeasy Tissue Kit (QIAGEN) eluting twice with 70 µl of EB buffer. A fragment (620 bp) of the mitochondrial cytochrome c oxidase subunit I gene (COI) was amplified for all individuals using the primers LCO1490 (5'-GGTCAACAAATCATAAAGATATTGG-3') and HCO2198 (5'-TAAACTTCAGGGTGACCAAAAAATCA-3') ([Folmer et al., 1994](#)). Individuals representing all COI haplotypes found were sampled to amplify a fragment of 16S rRNA including some tRNA (16S, 759bp) with the primers: 16S-tRNA-Leu-Ala-Ser-Leu-LumbF2: 5'-CGA CTG TTT AAC AAA AAC ATT GC-3' and 16S-tRNA-Leu-Ala-Ser-Leu-LumbR2: 5'-GTT TAA ACC TGT GGC ACT ATT C -3' ([Perez-Losada et al., 2009](#)).

Reaction mixtures comprised: 4 µl PCR buffer 5X (PROMEGA), 2 µl 25 mM MgCl₂ (PROMEGA), 1 µl 10 mM dNTPs, 1 µl of each 10 µM primer, 1 U Taq DNA polymerase (PROMEGA), 1 µl genomic DNA, and sterile H₂O to a final volume of 20 µl. The PCR profile for COI was 94 °C (5 min), 35 cycles of [94 °C (30 s), 52 °C (30 s), 72 °C (1 min)], with a final extension of 10 min at 72 °C and for 16S, 95 °C (3 min), 35 cycles of [95 °C (30 s), 57 °C (1 min), 72 °C (1 min, 30 s)], with a final extension of 5 min at 72 °C.

PCR products were purified and sequenced by Eurofins (www.eurofinsgenomics.eu) using the same primers. Chromatograms were visualized in Sequencher v.5.0 (Gene Codes Corporation, Ann Arbor, MI, USA). Sequences are deposited in GenBank (accession numbers KP214557–KP214595).

2.3. Genetic data analyses

Sequences were aligned in CLUSTALX v. 2.0.12 ([Thompson et al., 1997](#)) using default settings. Haplotypes were retrieved in DNACollapser (FaBox 1.41) and a Bayesian phylogeny was estimated for each gene with MrBayes v. 3.2.2 ([Ronquist and Huelsenbeck, 2003](#)). GTR + I + G, the best-fit substitution model implemented in MrBayes, as indicated by jModelTest2 ([Darriba et al., 2012](#)), was used. Parameters in MrBayes were set to forty million generations and a frequency of sampling of 4,000, retrieving 10,000 trees. The analysis was run twice and 25% of the trees were discarded as burn-in. The remaining trees were combined to find the maximum *a posteriori* probability estimate for each clade. Sequences from the earthworm *Aporrectodea trapezoides* (JF313607; HQ621864), which belongs to a different family, Lumbricidae, were used as an out-group. We performed the same analysis for the COI and 16S datasets separately and concatenated. A COI haplotype network was constructed using TCS version 1.21 with statistical parsimony and a 50-step connection limit ([Clement et al., 2000](#)).

An ultrametric tree was generated for COI using BEAST v. 1.7.5 ([Drummond et al., 2012](#)) with the evolutionary rate calculated by [Chang et al. \(2008\)](#) for Megascolecidae (2.4 substitutions/MY). The analysis was conducted under a constant size coalescent model, an uncorrelated lognormal relaxed clock and by setting GTR + I + G as the evolutionary model. We performed 30 parallel runs, each of which included 10 million generations, sampling each 1,000th generation. We combined tree and log files in Logcombiner v.1.7 by resampling at a lower frequency (30,000) and visualized the results using Tracer v. 1.5 ([Rambaut and Drummond, 2007](#)). The final tree was generated by TreeAnnotator v.1.7 with a burnin of 2000.

Island lineages have been observed to increase the rate of non-synonymous substitutions in mitochondrial protein coding genes ([Johnson and Seger, 2001](#); [Woolfit and Bromham, 2005](#)). The percentage of non-synonymous substitutions was calculated after

translating COI sequences in DNAsp v 5.10.1 ([Librado and Rozas, 2009](#)) using the invertebrate mitochondrial code.

2.4. Environmental characterisation

Sampling sites were characterised by elemental analysis together with environmental variables, such as moisture, organic matter, texture and pH. Codes from 0 to 4 (five categories) were assigned for covering percentage of grass, moss, ferns and stones (0 = absence; 1 = 1–25%; 2 = 26–50%; 3 = 51–75%; 4 = 76–100%). Codes were assigned as well to the degree of human interference at the sites (0 = absence, natural areas; 1 = scarce, grasslands and groves; 2 = moderate, edges of unpaved roads or forests paths; 3 = high, foraged with grazing livestock, areas near houses; 4 = very high, active crops and gardens).

Soils were air-dried prior to return to the UK for analysis. In the UK, samples were again air-dried and sieved to <2 mm. Subsamples were oven dried at 105 °C and moisture content determined. Concentrations are expressed on an oven dried soil basis. For pH measurement 10 g of air-dried soil was added to 25 ml deionised water and shaken for 15 min; pH was then measured ([ISO, 2005](#)). Loss on ignition was used as a proxy for organic matter content. Oven dried soil was weighed into a crucible and ignited overnight at 500 °C in a muffle furnace after which mass loss was determined ([Rowell, 1994](#)). Bulk metal content was determined by aqua regia digestion ([Arnold et al., 2008](#)) followed by analysis using a Perkin Elmer Optima 7300 DV inductively coupled plasma optical emission spectrometry instrument. Detection limits ([Supplementary Table 1](#)) were calculated as the mean plus 6 times the standard deviation on ten replicate analyses of the blank calibration standard ([Gill, 1997](#)). Method blanks were run and results were blank corrected where appropriate (for Ca, Mg, Na and Zn). For quality control we digested NIST Certified reference material Soil SRM 2709 and compared our results to those obtained after a US EPA 3050 acid digest. Recoveries were between 90 and 110% for Ca, Cr, Cu, Fe, Mg, Mn, Na, Ni, Pb, Sr and Zn.

Soil particle size distributions were determined using a Malvern MasterSizer 2000 with a Hydro2000MU wet dispersion unit. One to two grammes of air-dried, <2 mm sieved-soil were analysed with an obscuration of between 5 and 25%. Accurate instrument performance was confirmed using a Malvern 15–150 µm quality audit standard and “general purpose sand, 40–100 mesh” purchased from Fisher. Rather than measuring soil moisture content in the field during sampling, water holding capacity of the soil samples was determined following the method in ISO guideline 11274 ([ISO, 1992](#)) to give an indication of the relative potential moisture content of the different samples.

2.5. Statistical analyses

All statistical analyses were performed in R version 3.1.1 ([R Development Core Team, 2014](#)), using stats, car, MASS, psych ([Revelle, 2011](#)) and vegan ([Oksanen et al., 2007](#)) libraries. We performed a preliminary analysis using simple correlations between the initial set of 31 variables (see [Supplementary Table 1](#)) and species and lineage abundances. We removed those variables not showing significant relationships and performed subsequent analyses with a set of 19. Principal component analysis (PCA) was then applied to describe the main sources of variation and relationships between the retained habitat features. The ‘varimax’ rotation method was used to increase the interpretation of axes and the number of PCA axes examined was determined by sequential eigenvalue reduction ([Legendre and Legendre, 1998](#)). The relationship between the abundance of each earthworm species or lineage as well as lineage diversity in a location was examined using

generalised linear models (GLM) with altitude, the abundance of the other earthworm species, and the two stressor gradients from PCA analysis (i.e. PC1 and PC2) as fixed factors. As previously validated in rivers (Maceda-Veiga et al., 2013) and highlighted by Körner (2007), altitude was used as a surrogate for the position of the sampling sites on the mountain, and summarised the role of natural spatial gradients in earthworm abundance. Log-transformation was applied to altitude and earthworm abundances to reduce heterogeneity of variances and increase model fitting. A quasi-poisson error distribution was used for abundances, whereas Gaussian errors were assumed for lineage diversity. Significance of explanatory variables (factor effects) in GLMs was assessed using a likelihood ratio test. Best models were selected using a manual stepwise backward deletion of non-significant terms from the full global models containing all four factors and interactions. Models were validated with q–q plots of residuals and plotting fitted vs. predicted values, and spatial correlation was examined using Durbin Watson tests. As the modelling approach employed lacks a true variation coefficient (i.e. R^2), we calculated a pseudo- R^2 coefficient as follows: (null deviance – residual deviance)/null deviance. To complement the results of GLMs and to test the robustness of our results, we performed a hierarchical partitioning analysis (“hier.part” function in R) (Walsh and Mac Nally, 2011) using the error distributions validated in the GLM approach. An advantage of this approach is that it controls for co-linearity amongst explanatory variables that, even at low levels, can cause variance inflation and lead to erroneous conclusions (Graham, 2003). Other modelling criteria such as AIC and model averaging are discouraged because co-linearity results in biased parameter estimates (Freckleton, 2011). We assessed the significance of HP models using a randomization test for hierarchical partitioning analysis (function “rand.hp” in R). Significance in HP analysis was based on the upper 0.95 confidence interval, but it was reached at $p < 0.05$ in the remaining statistical procedures. The mapping software SURFER® (Golden Software, Golden, Colorado) was used to convert the total metal load ($\text{mol} \cdot \text{kg}^{-1}$), altitude and earthworm abundance data sets into a series of 2D contour maps.

3. Results

3.1. Species and haplotype geographic distributions

Most of the sampled megascolecid that showed a wide distribution within São Miguel Island (Fig. 1B), belonged to *A. gracilis* ($n = 110$) or *A. corticis* ($n = 315$), whereas a small group of unclassified megascolecids ($n = 22$) occurred in four localities, and were dominant in only one. In total, 565 COI sequences were generated, including those samples received from the various global sources. Some localities were dominated by a single species or even a single haplotype in the case of *A. corticis*. The distribution of COI haplotypes for *A. corticis* is shown in Fig. 1C. Assignment of COI haplotypes is shown in Supplementary Tables 3 and 4 (Azorean and non-Azorean samples, respectively), distribution of species and lineages per site in Supplementary Table 1 and assignment of 16S haplotypes in Supplementary Table 5.

3.2. Haplotype network

The networks generated by TCS with the COI and 16S genes are shown in Fig. 2. For *A. gracilis* a common COI haplotype was found (HAP-1, $n = 152$), which was shared with non-Azorean specimens (Brazil, Malaysia, Mexico, Taiwan, USA and China). Some individuals from China, Taiwan and two populations in São Miguel showed COI haplotypes separated by a single nucleotide polymorphism (SNP) when compared to HAP-1, but they all shared a

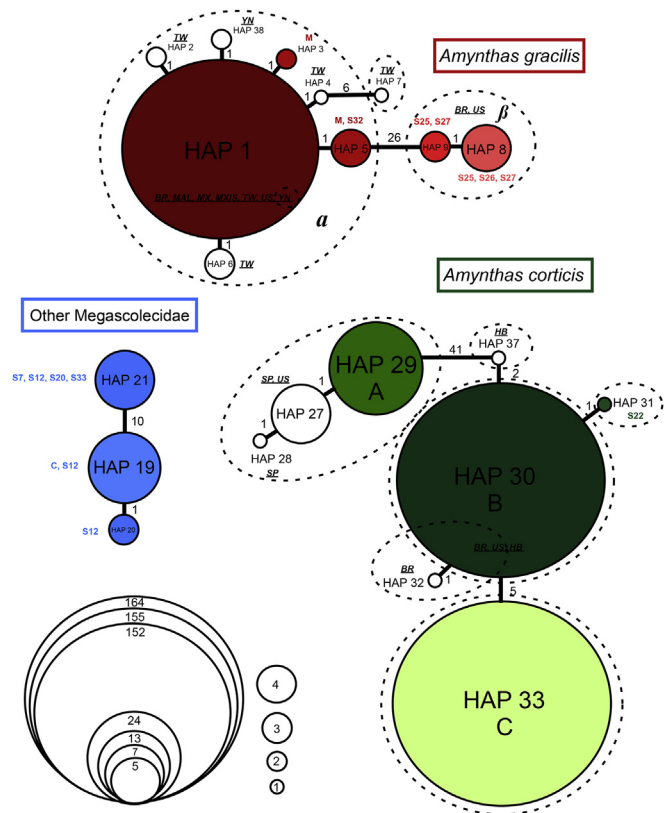


Fig. 2. COI network for all individuals. Coloured circles represent haplotypes present in the Azores; empty circles represent haplotypes present only outside the Azores. Red-brownish indicates *Amyntas gracilis* haplotypes; Green indicates *A. corticis* haplotypes. Blue indicates other Megascolecidae found in São Miguel. Names indicate origin of non-Azorean specimens (italics and underlined) or localities in São Miguel for the minority haplotypes. Geographic distribution of main haplotypes is found in Fig. 1. Size of the circles is proportional to the number of haplotypes. Number of SNPs separating haplotypes is shown on the connecting lines. Groups recovered using the less variable gene 16S are grouped within a dashed line. Only *Amyntas* individuals outside the Azores, and only in three instances (YN for *A. gracilis*; US and BR for *A. corticis*) showed different 16S haplotypes. See locality codes in Supplementary Tables 1 and 2. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

16S haplotype implying a common origin. This lineage was notated α , and was separated by 26 SNPs (0.043 uncorrected p-distance) from lineage β , composed of two COI haplotypes (HAP-8 and HAP-9) but sharing a 16S sequence. The last group comprised three *A. gracilis* populations from São Miguel (S25, S26, S27) and two non-Azorean specimens (USA and Brazil). The latter did not amplify for COI but showed the same 16S sequence.

For *A. corticis* three main haplogroups were found. Lineage A comprised 24 individuals from São Miguel with COI haplotype HAP-29 and specimens from Spain and USA with different COI haplotypes but sharing a 16S sequence. Lineage B included 155 individuals from the Azores, Brazil, USA, and China. Interestingly specimens from Brazil and USA showed a different 16S sequence (Fig. 2). Also a single individual at site S22 possessed different COI and 16S haplotypes. Lineages A and B were the most distinct (43 SNPs for COI, 0.069 uncorrected p-distance) but an intermediate haplotype from a Chinese sample (HAP-37) was found. Finally, lineage C possessed only 5 SNPs different to lineage B (i.e. 0.008 COI uncorrected p-distance) and comprised only individuals collected from the Azores ($n = 164$). Lineage B seems to be the origin of the other groups according to TCS, although intermediates may be missing from our sampling.

3.3. Phylogenetic trees and dating

The topologies of the phylogenetic trees were consistent for all analyses (separate or concatenated datasets; Fig. 3) and provided concordant groupings with the network analyses. Dates recovered

using BEAST imply that the divergences between the identified lineages are very old and certainly pre-dated human introductory events. For *A. gracilis*, the α and β split was dated to around 1.14 Mya. For *A. corticis*, lineage A was dated as having diverged at 1.93 Mya from B and C, with the latter pair diverging approximately 267 Kya.

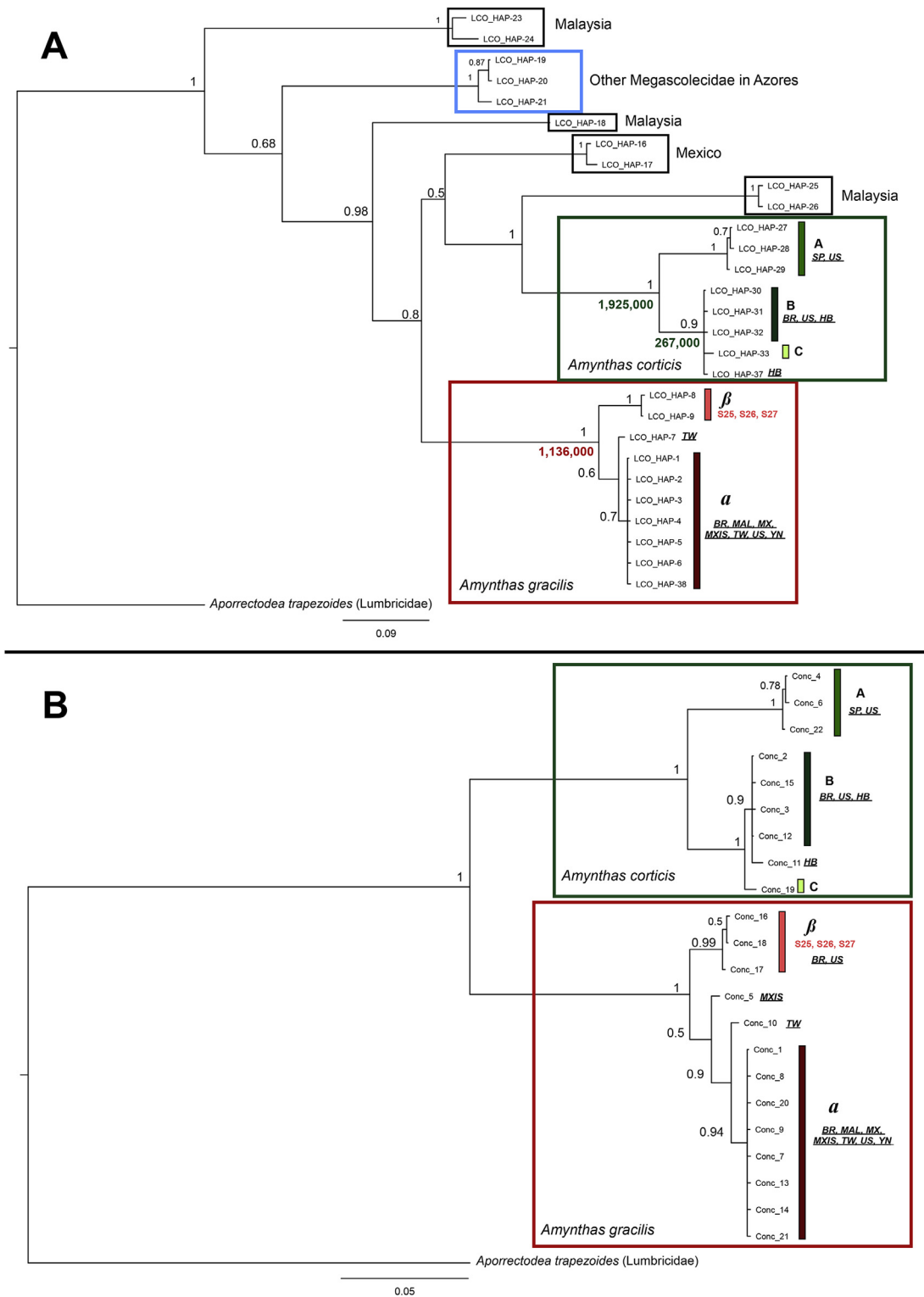


Fig. 3. Bayesian phylogenies for the mitochondrial markers analysed for the *Amynthes* dataset. A: COI gene – posterior probabilities (above) and the dates generated by BEAST for relevant ancestors originating the defined groups (below) are shown on the nodes. B: concatenated data set, including only *A. gracilis* and *A. corticis*. Posterior probabilities are shown on the nodes. Clades identified with a vertical bar represent haplotypes present in Azores. Codes for non-Azorean specimens are shown in italics and underlined. See locality codes in Supplementary Tables 1 and 2 and haplotype codes in Supplementary Tables 4, 5 and 7.

3.4. Non-synonymous substitutions

Relatively low numbers of non-synonymous substitutions were generally found between pairs of sequences ranging from 0 to 16.7%, even when comparing the different megascolecid species analysed using up to 126 substitutions. However, percentages of non-synonymous substitutions between 28.6 and 100% were found between *A. corticis* haplotypes involving the singly observed haplotypes 28, 31, 32 and 37. Nevertheless it is worth highlighting that between the *A. corticis* haplotypes 30 and 33, which seem to be well-established because of their geographical distribution and abundances, we found 40% non-synonymous substitutions (two out of five).

3.5. Relationship between earthworm abundance and environmental variables

Elemental concentrations and habitat features were summarised using PCA, producing two significant axes explaining 72% of the total variation (Table 1). PC1 was found to be loaded on element concentrations and accounted for 52% of the variation, whereas PC2 included habitat features, representing the degree of human influence, and explained 20% of the variation. As the environmental gradients built with the PCA and the abundance of earthworms were significantly correlated with altitude (Spearman's rho range = 0.3–0.7), it was necessary to disentangle this relationship and analyse the independent and joint effects of these predictors on the abundance of each species of earthworm or clade groups. The results of GLM and HP models were mostly concordant indicating that co-linearity between predictors played a minor role in our data-set. Discrepancies were only observed between GLM and HP when the former retained as significant the PC2 gradient due to its highly significant and positive relationship with altitude ($r = 0.7$, $p < 0.01$). None of the GLMs exhibited spatial autocorrelation (i.e. dependency amongst observations in a geographic space) in the residuals according to Durbin Watson tests ($D-W > 1.7$, All $p > 0.10$).

At least one of the environmental gradients from the PCA and/or the abundance of the other earthworm species was retained as significant, and made the largest contribution to the abundance of the earthworm species included as a response variable (Tables 2 and 3). Interestingly, the abundance of *A. gracilis* was negatively associated with the abundance of *A. corticis* (-1.43 ± 0.43 ;

Table 1

Loadings for axes 1 and 2 according to PCA built with soil characteristics and other habitat features measured in São Miguel Island (Azores). Bold values are considered high ≥ 0.7 .

Environmental variables	PC1	PC2
Ba	0.93	0.10
Ca	0.94	0.16
Cr	0.93	0.24
Cu	0.77	0.32
Fe	0.90	0.23
Ga	0.95	0.06
K	0.43	0.39
Mg	0.96	0.13
Mn	0.37	0.63
Na	0.73	0.10
Ni	0.93	0.18
Sr	0.92	0.13
Ti	0.96	0.07
Zn	0.40	0.65
Soil pH	0.61	0.58
Ferns cover	−0.07	−0.51
Moss cover	−0.12	− 0.90
Stones cover	0.03	−0.50
Human influence	−0.02	0.91

Table 2

Results of the final GLM models for the abundance of each earthworm species and lineage, and the lineage diversity that include the significant variables highlighted in previous full models with interactions. Pseudo- R^2 indicates the proportion of variation explained by the model (see Methods).

Response variable	Explanatory variables	Estimate \pm SE	df	χ^2	p-value	Pseudo- R^2 (%)
<i>A. gracilis</i>	<i>A. corticis</i>	-1.43 ± 0.43	1,30	10.72	<0.001	24.2
Lineage α	<i>A. corticis</i>	-1.72 ± 0.49	1,30	12.51	<0.001	30.1
Lineage β	Metal concentrations (PC1)	-3.18 ± 1.51	1,29	15.53	<0.001	22.6
	Human influence (PC2)	1.56 ± 0.65	1,29	8.12	0.004	
<i>A. corticis</i>	Metal concentrations (PC1)	-0.20 ± 0.09	1,29	5.84	0.015	32.6
Lineage A	<i>A. gracilis</i>	-0.69 ± 0.21	1,29	13.07	<0.001	
	Human influence (PC2)	1.78 ± 0.70	1,30	10.73	<0.001	38.1
Lineage B	<i>A. gracilis</i>	-0.75 ± 0.38	1,30	4.52	0.03	10.4
Lineage C	Altitude	1.97 ± 0.53	1,29	15.22	<0.001	43.8
	Metal concentrations (PC1)	-0.31 ± 0.15	1,29	4.61	0.03	
Lineage diversity	Metal concentrations (PC1)	-0.48 ± 0.16	1,30	8.08	0.004	21.2

$\chi^2 = 10.72$, $df = 1$, $P < 0.001$), suggesting spatial segregation/competition between the two invasive species (Fig. 4). The abundance of *A. gracilis* also made the largest contribution to explain the abundance of *A. corticis* (-0.69 ± 0.21 ; $\chi^2 = 13.06$, $df = 1$, $P < 0.001$) followed by the metal concentrations (PC1) (-0.20 ± 0.02 ; $\chi^2 = 5.84$, $df = 1$, $P = 0.02$). When the abundance of each lineage was considered in the analysis, metal concentration was also identified as an important factor explaining the abundance of *A. corticis* from lineage C and *A. gracilis* from lineage β (Table 2). *A. corticis* lineage C worms were, however, mostly associated with higher altitude (1.96 ± 0.56 ; $\chi^2 = 15.22$, $df = 1$, $P < 0.001$) and according to HP, with localities with a high degree of habitat naturalness (PC2) (Table 3, Fig. 5). The opposite relationship was found between habitat naturalness and the abundance of *A. corticis* from lineage A (1.78 ± 0.70 ; $\chi^2 = 10.73$, $df = 1$, $P = 0.001$) and *A. gracilis* from lineage β (1.56 ± 0.65 ; $\chi^2 = 8.12$, $df = 1$, $P = 0.004$), both being positively related with human influence. This illustrates that habitat requirements may differ between lineages from the same invasive species. Finally, we found that lineage diversity was affected negatively by the metal load (PC1) (Table 2).

Table 3

Independent contribution (%) of the environmental predictors and the abundance of the other invasive species to the explained variation of the hierarchical partitioning models performed on each earthworm species and lineage, and the lineage diversity. Bold values indicated the highest independent contribution that achieved significance (*). Significance was reached at the 95% confidence interval based on a randomized permutation test (rand.hp function, see Methods).

Response variable	Altitude	Metal concentration (PC1)	Human influence (PC2)	Other worm species
<i>A. gracilis</i>	3.70	3.87	3.61	29.74
Lineage α	3.21	1.60	1.73	32.10
Lineage β	1.41	14.50	7.75	0.60
<i>A. corticis</i>	2.69	13.93	4.84	26.79
Lineage A	4.76	0.54	15.37	3.7
Lineage B	1.57	2.60	0.30	13.40
Lineage C	22.88	16.30	14.01	3.78
Lineage diversity	2.00	19.00	3.00	–

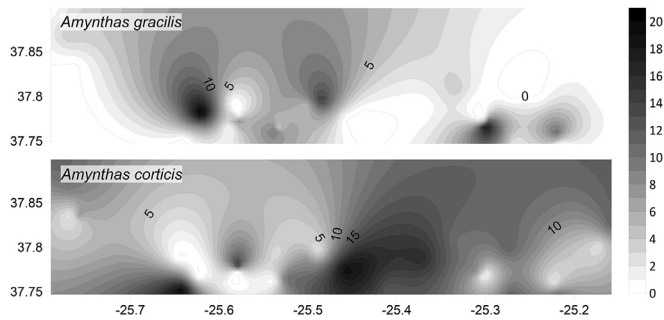


Fig. 4. Surface maps generated with SURFER® showing the overlaid levels of abundance of individuals of *Amyntas gracilis*, and *A. corticis* derived from 32 collection sites in São Miguel Island, Azores. Geographical coordinates in decimal degrees are represented by x and y and the location of the corresponding sampling points are shown in Supplementary Table 1 in the same units.

4. Discussion

Our results show that *Amyntas* species are successful invaders of São Miguel Island. They appear to be present in all habitable soils across the island. Lineage analysis based on mitochondrial genes, and compared with the genotypes of non-Azorean populations, strongly indicates that the extant populations result from multiple introductions. The distributions of the two main megascolecid species *A. gracilis* and *A. corticis* are negatively correlated, indicating spatial segregation of the invaders. *A. corticis* seems to be more widespread and therefore more successful in occupying the diverse habitats found on the island; this was particularly the case with *A. corticis* lineage C, which was the only lineage which we were unable to identify in any non-Azorean population. We found that besides the negative correlation among the two invasive species, altitude, soil parameters and habitat were also linked to the distribution patterns of the different mitochondrial lineages and to lineage diversity.

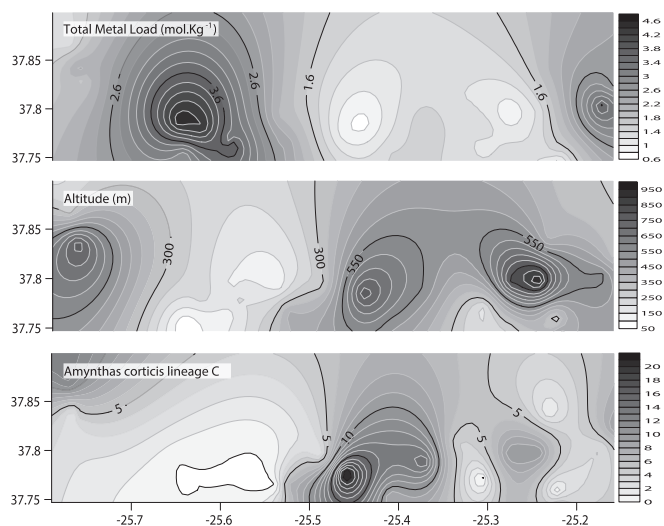


Fig. 5. Surface maps generated with SURFER® showing the overlaid levels of abundance of individuals of *Amyntas corticis* lineage C, total metal load in soils ($\text{mol} \cdot \text{kg}^{-1}$), and altitude (m) derived from 32 collection sites in São Miguel island, Azores. Geographical coordinates in decimal degrees are represented by x and y and the location of the corresponding sampling points are shown in Supplementary Table 1 in the same units.

4.1. *Amyntas* species as successful invaders of São Miguel Island

Amyntas species are widely distributed across São Miguel as shown by our widespread sampling. Earthworms of this genus have previously shown high invasive potential, and are now widely spread over tropical and temperate geographic regions around the globe although with particular habitat preferences mainly associated with anthropogenic disturbance (Hendrix et al., 2008). Different characteristics of certain earthworm species have been noted to contribute to successful invasion, such as the ability to aestivate, physiological plasticity, and parthenogenesis (Lavelle et al., 1999; Edwards, 2004). *Amyntas* could be particularly aided by its high mobility and reproduction rate (Burtelov et al., 1998; Garcia and Fragoso, 2002), and parthenogenesis in the case of *A. corticis* (see Section 4.3). They are also epigeic species (surface dwellers), which disperse actively through the soil but are also prone to physical dispersal forces such as flooding, wind or phoresy (Eijsackers, 2011). Success of *Amyntas* species as invaders in the particular setting of São Miguel may be due to several factors identified here. Firstly, multiple introduction events could promote adaptation to the new environment by enhancing genetic diversity (Cameron et al., 2008). Such events would counteract the genetic bottleneck resulting from a singular introduction (Caron et al., 2013). We have inferred at least four distinct *Amyntas* introduction events in São Miguel, two for each species (i.e. *A. corticis* A, B; *A. gracilis* α , β). Lineage C in *A. corticis* remained of unknown origin (see below). More COI haplotypes were found but 16S clustered them into those five groups. The number of times that a species arrives has been listed as an important factor for a successful colonizer, together with endurance of environmental conditions and the range of habitats in which a species can establish (Eijsackers, 2011). Habitat matching facilitates invasion (Hendrix et al., 2006). As stated by Beddard (1912), temperate species tend to invade temperate regions and tropical species tend to invade tropical regions. Consequently, the similarity of the Azores sub-tropical climate to that of the *Amyntas* source-regions in Asia has likely benefitted the worms, enabling the pioneers to survive and subsequently establish sustainable populations. Also, being epigeic species they are expected to have found the shallow but relatively fertile Azorean volcanic soils, with ample and widespread associated surface litter (pers. obs.), compatible with their ecological requirements.

4.2. Negative correlation of abundances of both invaders

The negative relationship between the abundances of *A. gracilis* and *A. corticis* was found in this study to make the largest independent contribution to explain their respective abundances in São Miguel. Many studies have analysed the effects of invasive species on native relatives, including niche displacement or competitive exclusion (e.g., Mooney and Cleland, 2001; Johnson et al., 2009), although this effect is not as clear in earthworms (Hendrix et al., 2006; González et al., 2006). However, the interaction between two invasive species from the same taxon has not been studied extensively, although some studies of multiple invaders from different animal groups provide evidence of facilitation (meltdown hypothesis) among invasive species (Ricciardi, 2001; O'Dowd et al., 2003; Grosholz, 2005), and only a few show negative interactions (Ross et al., 2004; Johnson et al., 2009). Other studies have shown spatial segregation of introduced related species, indicating a similar use of resources by both (Allen and Shea, 2006; Rauschert, 2006; Monroy et al., 2014). Hitherto, the laboratory-based study on lumbricids by Butt (1998) appears to be the only one that has examined interactions between potential invasive earthworm species; he concluded that the interaction showed the hallmarks of

facilitation. To our knowledge the present field-based study of two closely related megascoleids is the first record of a negative correlation in the abundances of the invaders. We cannot discount the possibility of an initial facilitation between the species, but their similar resource requirements along with different adaptation capabilities (see Section 4.4) may have caused their subsequent exclusion. Uvarov (2009) concluded that, unlike deep-burrowing anecic earthworms, which mainly show positive interactions, epigeic earthworms are mostly competitive, and especially so in the acquisition of food (Abbott, 1980). However, the extent of competition among *Amyntas* species in São Miguel is uncertain, and possibly confounded by edaphic preferences, because they have been found together in some sites. Richard et al. (2012) postulated that interspecific interactions are important factors driving the spatial distribution of earthworms at a local scale. Future studies on the distribution of the earthworm fauna of São Miguel Island could possibly benefit from focusing a more highly resolved spatial scale in order to shed light on the inter-play between interspecific and abiotic environmental parameters.

4.3. *A. corticis* as a more successful colonizer than *A. gracilis*

A. corticis showed a broader distribution across São Miguel Island and has colonised a wider variety of habitats than *A. gracilis*. Our results suggest that the different lineages of *A. corticis* have colonised different soil types (see Section 4.4), which may indicate higher physiological plasticity and adaptation potential. *A. corticis* is recognized to be widely tolerant to environmental factors (Fragoso et al., 1999). Between the two species, *A. corticis* has greater mobility (*pers. observ.*) and also reproduces parthenogenetically in the Azores, unlike the sexually reproducing *A. gracilis* (author's work in progress). Parthenogenetic earthworms, most of them also known to be polyploid (Muldal, 1952; Jaenike and Selander, 1979; Viktorov, 1997; Lavelle et al., 1999; Terhivuo and Saura, 2006), have several fitness advantages that would promote colonization and an ability to sustain stable populations in new environments since a single organism can establish an entire population. Higher levels of genetic variability and exceptionally plastic genomes that permit rapid adaptation, supported by higher reproductive rates enables the production of a disproportionate number of offspring that, in turn, repeat the parental dispersion behaviour and reproductive performance, thus extending and sustaining niche occupation (Hughes, 1989; Terhivuo and Saura, 2006; Díaz Cosín et al., 2011). As such, these differences in biology between the two species may contribute to the relative success of *A. corticis* in colonising new habitats.

4.4. Environmental requirements of different lineages

Individuals belonging to the five main mitochondrial lineages identified for *A. gracilis* and *A. corticis* have different habitat requirements that would have been overlooked by the analysis at species level. These results stress the importance of the genetic characterization of invasive species to better understand their invasion patterns. Other studies have also found that genetic lineages of the same or related species have different ecological requirements (VanDyke et al., 2004; Mozdzer and Zieman, 2010; Gul, 2013). Such genetically-linked differences probably modulate their invasive potential. In the current study, metal concentration in soil, as broadly defined by PC1, was negatively associated with the abundance of lineage C in *A. corticis* and β in *A. gracilis*. Soil features, such as texture and elemental composition, seem to be more important for colonization by earthworms than inherent ecological characteristics (such as *r* and *K* strategies) and they also affect dispersal (Eijsackers, 2011). Invasive earthworms can avoid adverse

conditions, such as extremely acidic, extremely alkaline or contaminated soils (Eijsackers, 2011). In the current study altitude was a significant determinant of the abundance of lineage C of *A. corticis*. A positive effect of altitude on this lineage, plus its negative relationship with human influence and soil metal concentrations, seems to indicate that this lineage is related to the most natural conditions in the island. Suarez et al. (2006) showed that exotic earthworms were more likely to occur on low and flat sites in comparison to more remote, steep sites; lineage C of *A. corticis* may be an exception that proves the rule. However, according to the geographic parthenogenesis theory (Vandel, 1928), parthenogens can colonize higher altitudes than their non-parthenogenetic conspecifics. They also have broader distribution areas (Cosendai et al., 2013 and references therein), which could explain why at least one lineage of *A. corticis* inhabits higher altitudes than other *A. corticis* lineages and each of the *A. gracilis* lineages.

Species invasions are frequently underpinned by anthropogenic drivers and hence the establishment of invasive species may frequently be related to measures of human influence. (Hendrix et al., 2008). Abundance of lineage β of *A. gracilis* and lineage A of *A. corticis* showed a positive relationship with the degree of human influence supporting this statement only for those lineages. Habitats altered by humans are also characterized by increased species richness and dominance by alien species (Borges et al., 2006). We found no relationship between the number of lineages present at a site and the degree of human influence but our results show that more lineages were present in places with lower metal concentrations.

The fact that different lineages can tolerate different conditions could positively influence the success of the invasion of *Amyntas* in the Azores. Thus representatives of the two species and their different lineages appear to inhabit the full range of available volcanic soil types on São Miguel Island.

4.5. Lineage C of *A. corticis* has no known origin

All the haplotypes, except for lineage C (haplotype 33 for COI) in *A. corticis*, could be detected outside the Azores and therefore must have been introduced independently to the island, having diverged beforehand. Another exception was haplotype 31, but it was found in only one individual. Lineage C is the most widespread in São Miguel and is related to the most natural conditions in the island (see above). It could be argued that lineage C has evolved within the island and is thus related to the island's natural environmental characteristics. In fact, the percentage of non-synonymous substitutions of COI between lineages B and C (haplotype 30 and haplotype 33) of *A. corticis* in comparison with the remainder pairwise values, supports an island origin for the latter lineage. It has been shown that island lineages have increased rates of non-synonymous substitutions in mitochondrial protein coding genes, apparently starting soon after the isolation (Johnson and Seger, 2001). This phenomenon contradicts the dates given by the calibrated tree, which indicates an ancient separation of these two lineages and, therefore, a high probability that lineage C evolved outside the Azores and was subsequently introduced. It is conceivable that the tree dates could be valid for natural colonization, but such an event seems improbable due to the oceanic isolation of the entire Azorean archipelago (but see Sharma and Giribet, 2012). However evolutionary rates cannot always be extrapolated from one group to another since different rates occur even within the same family (Novo et al., 2012). Also, rapid evolution has been proposed in island environments (Aleixandre et al., 2013), even over relatively short time scales from a few decades up to several thousands of years (Millien, 2006). This could justify the

old dates in the tree for the split between lineages B and C when applying non-specific rates. Other studies show the lack of evidence of an increase in evolutionary rates in island taxa (Bromham and Woolfit, 2004), but still demonstrate the existence of higher ratios of non-synonymous to synonymous substitutions (Woolfit and Bromham, 2005). It is prudent to conclude that neither the evolution of lineage C within the island, nor evolution outside the island and coupled with posterior human-mediated introduction, can be discarded after this study. Wider collection in the home range of the species may be useful to try and discover the existence of lineage C outside the island.

4.6. Conclusion

The studied *Amyntas* species are successful invaders in the Azores due to their inherent biological and ecological characteristics and enhanced by the fact that their presence is the result of multiple introductions. The relatively higher success of *A. corticis* may be due to parthenogenesis and higher plasticity. One of its lineages has an unknown origin and seems to inhabit more natural, undisturbed, areas on the island. Abundances of *A. gracilis* and *A. corticis* are negatively correlated, suggesting segregation or competition between both invaders as opposed to the 'meltdown' hypothesis. Distinct genetic lineages showed different environmental preferences, with metal concentrations, degree of human influence and altitude linked to their abundances. Lineage diversity was higher in places with lower metal concentrations. Our results highlight the need for genetic studies in order to better understand invasion patterns since a single species can present complex responses to new environments. The inclusion of nuclear markers such as microsatellites or RAD-tag analysis of *Amyntas* species found in the Azores populations could further develop understanding about the more recent adaptation mechanisms to further explain observed distribution patterns and possible competition of *Amyntas* species in São Miguel. Finally, we envisage that a suite of genotyping and bioinformatics approaches such as those used in the present study, combined with the mentioned nuclear markers, could be invaluable tools to help characterise and manage invasive species in diverse aquatic as well as terrestrial ecosystems worldwide.

Acknowledgements

We would like to thank George Brown, Emma Sherlock, Carlos Fragoso, Chih-Han Chang, Isabelle Barois, Nor Azwady Abd, Darío Díaz Cosín, Jiun-Hong Chen, Jibao Jiang, Jin Sun, Qi Zhao and Prof. Qiu for their kindness and donation of samples for this study and Samuel James and Jose Antonio Garcia Perez for their useful comments on *Amyntas* biology. MN was supported by a postdoctoral Fellowship from the Spanish Government and a Marie Curie Fellowship (EpiWorm: FP7-PEOPLE-2012-329690). AMV was funded by a Marie Curie Fellowship (Para-Tox: FP7-PEOPLE-2012-327941). This study was funded by the grant NERC NE/I026022/1.

Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.soilbio.2015.02.031>.

References

Abbott, I., 1980. Do earthworms compete for food. *Soil Biology & Biochemistry* 12, 523–530.

Alexandre, P., Montoya, J.H., Mila, B., 2013. Speciation on oceanic islands: rapid adaptive divergence vs. cryptic speciation in a Guadalupe Island Songbird (*Aves: Junco*). *Plos One* 8.

Allen, M.R., Shea, K., 2006. Spatial segregation of congeneric invaders in central Pennsylvania, USA. *Biological Invasions* 8, 509–521.

Amorim, I.R., Emerson, B.C., Borges, P.A.V., Wayne, R.K., 2012. Phylogeography and molecular phylogeny of Macaronesian island Taphius (Coleoptera: Zopheridae): why are there so few species in the Azores? *Journal of Biogeography* 39, 1583–1595.

Arnold, B.E., Hodson, M.E., Charnock, J., Peijnenburg, W.J.G.M., 2008. Comparison of subcellular partitioning, distribution, and internal speciation of Cu between Cu-tolerant and naive populations of *Dendrodrilus rubidus* Savigny. *Environmental Science & Technology* 42, 3900–3905.

Baker, H.G., 1965. Characteristics and modes of origin of weeds. In: Baker, H.G., Stebbins, G.L. (Eds.), *The Genetics of Colonizing Species*. Academic Press, New York, pp. 147–168.

Baker, H.G., 1967. Support for Baker's law-as a rule. *Evolution* 21, 853–856.

Beddard, F.E., 1912. *Earthworms and Their Allies*. Univ. Press, Cambridge, UK.

Bohlen, P.J., Groffman, P.M., Fahey, T.J., Fisk, M.C., Suarez, E., Pelletier, D.M., Fahey, R.T., 2004. Ecosystem consequences of exotic earthworm invasion of north temperate forests. *Ecosystems* 7, 1–12.

Borges, P.A.V., Lobo, J.M., de Azevedo, E.B., Gaspar, C.S., Melo, C., Nunes, L.V., 2006. Invasibility and species richness of island endemic arthropods: a general model of endemic vs. exotic species. *Journal of Biogeography* 33, 169–187.

Bromham, L., Woolfit, M., 2004. Explosive radiations and the reliability of molecular clocks: island endemic radiations as a test case. *Systematic Biology* 53, 758–766.

Buckley, T.R., James, S., Allwood, J., Bartlam, S., Howitt, R., Prada, D., 2011. Phylogenetic analysis of New Zealand earthworms (Oligochaeta: Megascotocidae) reveals ancient clades and cryptic taxonomic diversity. *Molecular Phylogenetics and Evolution* 58, 85–96.

Burtelew, A.E., Bohlen, P.J., Groffman, P.M., 1998. Influence of exotic earthworm invasion on soil organic matter, microbial biomass and denitrification potential in forest soils of the northeastern United States. *Applied Soil Ecology* 9, 197–202.

Butt, K.R., 1998. Interactions between selected, earthworm species: a preliminary, laboratory-based study. *Applied Soil Ecology* 9, 75–79.

Cameron, E.K., Bayne, E.M., Coltman, D.W., 2008. Genetic structure of invasive earthworms *Dendrobaena octaedra* in the boreal forest of Alberta: insights into introduction mechanisms. *Molecular Ecology* 17, 1189–1197.

Carine, M.A., Schaefer, H., 2010. The Azores diversity enigma: why are there so few Azorean endemic flowering plants and why are they so widespread? *Journal of Biogeography* 37, 77–89.

Caron, V., Norgate, M., Ede, F.J., Nyman, T., Sunnucks, P., 2013. Novel microsatellite DNA markers indicate strict parthenogenesis and few genotypes in the invasive willow sawfly *Nematus oligospilus*. *Bulletin of Entomological Research* 103, 74–88.

Chang, C.H., Chen, J.H., 2005. Taxonomic status and intraspecific phylogeography of two sibling species of *Metaphire* (Oligochaeta: Megascotocidae) in Taiwan. *Pedobiologia* 49, 591–600.

Chang, C.H., Lin, S.M., Chen, J.H., 2008. Molecular systematics and phylogeography of the gigantic earthworms of the *Metaphire formosae* species group (Clitellata, Megascotocidae). *Molecular Phylogenetics and Evolution* 49, 958–968.

Clement, M., Posada, D., Crandall, K.A., 2000. TCS: a computer program to estimate gene genealogies. *Molecular Ecology* 9, 1657–1659.

Cosendai, A.C., Wagner, J., Ladinig, U., Rosche, C., Horandl, E., 2013. Geographical parthenogenesis and population genetic structure in the alpine species *Ranunculus kuepferi* (Ranunculaceae). *Heredity* 110, 560–569.

Cunha, L., Montiel, R., Novo, M., Orozco-terWengel, P., Rodrigues, A., Morgan, A.J., Kille, P., 2014. Living on a volcano's edge: genetic isolation of an extremophile terrestrial metazoan. *Heredity* 112, 132–142.

Darriba, D., Taboada, G.L., Doallo, R., Posada, D., 2012. jModelTest 2: more models, new heuristics and parallel computing. *Nature Methods* 9, 772.

Díaz Cosín, D.J., Novo, M., Fernández, R., 2011. Reproduction of earthworms: sexual selection and parthenogenesis. In: Karaka, A. (Ed.), *Biology of Earthworms*. Springer, Berlin, pp. 69–86.

Drummond, A.J., Suchard, M.A., Xie, D., Rambaut, A., 2012. Bayesian Phylogenetics with BEAUTi and the BEAST 1.7. *Molecular Biology and Evolution* 29, 1969–1973.

Edwards, C.A. (Ed.), 2004. *Earthworm Ecology*. CRC Press, Boca Raton, FL.

Eijssackers, H., 2011. Earthworms as colonizers of natural and cultivated soil environments. *Applied Soil Ecology* 50, 1–13.

Fernandez, R., Almodovar, A., Novo, M., Gutierrez, M., Cosin, D.J.D., 2011. A vagrant clone in a peregrine species: phylogeography, high clonal diversity and geographical distribution in the earthworm *Aporrectodea trapezoides* (Duges, 1828). *Soil Biology & Biochemistry* 43, 2085–2093.

Folmer, O., Black, M., Hoeh, W., Lutz, R., Vrijenhoek, R., 1994. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* 3, 294–299.

Fragoso, C., Kanyonyo, J., Moreno, A., Senapati, B.K., Blanchart, E., Rodríguez, C., 1999. A survey of tropical earthworms: taxonomy, biogeography and environmental plasticity. In: Lavelle, P., Brissard, L., Hendrix, P.F. (Eds.), *Earthworm Management in Tropical Agroecosystems*. CAB International, Wallingford, U.K, pp. 1–26.

Freckleton, R.P., 2011. Dealing with collinearity in behavioural and ecological data: model averaging and the problems of measurement error. *Behavioral Ecology and Sociobiology* 65, 91–101.

- Frelich, L.E., Hale, C.M., Scheu, S., Holdsworth, A.R., Heneghan, L., Bohlen, P.J., Reich, P.B., 2006. Earthworm invasion into previously earthworm-free temperate and boreal forests. *Biological Invasions* 8, 1235–1245.
- García, J.A., Fragoso, C., 2002. Growth, reproduction and activity of earthworms in degraded and amended tropical open mined soils: laboratory assays. *Applied Soil Ecology* 20, 43–56.
- Gill, R., 1997. *Modern Analytical Geochemistry*. Longman, Harlow, UK.
- Gomes, A., Gaspar, J.L., Queiroz, G., 2006. Seismic vulnerability of dwellings at Sete Cidades Volcano (S. Miguel Island, Azores). *Natural Hazards and Earth System Sciences* 6, 41–48.
- Gonzalez, G., Huang, C.Y., Zou, X.M., Rodriguez, C., 2006. Earthworm invasions in the tropics. *Biological Invasions* 8, 1247–1256.
- Graham, M.H., 2003. Confronting multicollinearity in ecological multiple regression. *Ecology* 84, 2809–2815.
- Grosholz, E.D., 2005. Recent biological invasion may hasten invasional meltdown by accelerating historical introductions. *Proceedings of the National Academy of Sciences of the United States of America* 102, 1088–1091.
- Gul, S., 2013. Ecological divergence between two evolutionary lineages of *Hyla savignyi* (Audouin, 1827) in Turkey: effects of the Anatolian Diagonal. *Animal Biology* 63, 285–295.
- Hale, C.M., Frelich, L.E., Reich, P.B., 2005. Exotic European earthworm invasion dynamics in northern hardwood forests of Minnesota, USA. *Ecological Applications* 15, 848–860.
- Harris, D.J., Ferreira, A.F., Martins, A.M.D., 2013. High levels of mitochondrial DNA diversity within oxychilid land snails (subgenus *Drouetia* Gude, 1911) from São Miguel island, Azores. *Journal of Molluscan Studies* 79, 177–182.
- Hendrix, P.F., Baker, G.H., Callahan, M.A., Damoff, G.A., Fragoso, C., Gonzalez, G., James, S.W., Lachnicht, S.L., Winsome, T., Zou, X., 2006. Invasion of exotic earthworms into ecosystems inhabited by native earthworms. *Biological Invasions* 8, 1287–1300.
- Hendrix, P.F., Callahan, M.A., Drake, J.M., Huang, C.Y., James, S.W., Snyder, B.A., Zhang, W.X., 2008. Pandora's box contained bait: the global problem of introduced earthworms. *Annual Review of Ecology Evolution and Systematics* 39, 593–613.
- Hernández, P., Fernández, R., Novo, M., Trigo, D., Cosín, D.J.D., 2007. Geostatistical and multivariate analysis of the horizontal distribution of an earthworm community in El Molar (Madrid, Spain). *Pedobiologia* 51, 13–21.
- Hughes, R.N., 1989. *A Functional Biology of Clonal Animals*. Chapman and Hall, London; New York xii, 331 pp.
- ISO, 2005. Determination of pH. Switzerland, Geneva.
- ISO, 1992. Soil quality e determination of water retention characteristics. Laboratory Methods. No. 11274. Geneva.
- Jaenike, J., Selander, R.K., 1979. Evolution and ecology of parthenogenesis in earthworms. *American Zoologist* 19, 729–737.
- Jamieson, B.G.M., Tillier, S., Tillier, A., Justine, J.L., Ling, E., James, S., MacDonald, K., Huggall, A.F., 2002. Phylogeny of the Megascolecidae and Crassicitellata (Annelida, Oligochaeta): combined versus partitioned analysis using nuclear (28S) and mitochondrial (12S, 16S) rDNA. *Zoosystema* 24, 707–734.
- Jiménez, J.J., Decaens, T., Lavelle, P., Rossi, J.P., 2014. Dissecting the multi-scale spatial relationship of earthworm assemblages with soil environmental variability. *BMC Ecology* 14, 26.
- Johnson, K.P., Seger, J., 2001. Elevated rates of nonsynonymous substitution in island birds. *Molecular Biology and Evolution* 18, 874–881.
- Johnson, P.T.J., Olden, J.D., Solomon, C.T., Vander Zanden, M.J., 2009. Interactions among invaders: community and ecosystem effects of multiple invasive species in an experimental aquatic system. *Oecologia* 159, 161–170.
- Körner, C., 2007. The use of 'altitude' in ecological research. *Trends in Ecology and Evolution* 22, 569–574.
- Lavelle, P., Brussaard, L., Hendrix, P.F., 1999. *Earthworm management in tropical agroecosystems*. CABI Pub. USA, xii, Wallingford, Oxon, UK ; New York, NY, 300 pp.
- Legendre, P., Legendre, L., 1998. *Numerical Ecology*. second English ed.. Elsevier, Amsterdam ; New York. xv, 853 pp.
- Librado, P., Rozas, J., 2009. DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* 25, 1451–1452.
- Louvat, P., Allègre, C.J., 1998. Riverine erosion rates on São Miguel volcanic island, Azores archipelago. *Chemical Geology* 148, 177–200.
- Maceda-veiga, A., De Sostoa, A., Sánchez-Espada, S., 2013. Factors affecting the establishment of the invasive crayfish *Procambarus clarkii* (Crustacea, Decapoda) in the Mediterranean rivers of the northeastern Iberian Peninsula. *Hydrobiologia* 703, 33–45.
- Malucelli, F., Terribile, F., Colombo, C., 1999. Mineralogy, micromorphology and chemical analysis of andosols on the Island of São Miguel (Azores). *Geoderma* 88, 73–98.
- Millien, V., 2006. Morphological evolution is accelerated among island mammals. *Plos Biology* 4, 1863–1868.
- Monroy, M., Maceda-veiga, A., Caiola, N., De Sostoa, A., 2014. Trophic interactions between native and introduced fish species in a littoral fish community. *Journal of Fish Biology* 85, 1693–1706.
- Mooney, H.A., Cleland, E.E., 2001. The evolutionary impact of invasive species. *Proceedings of the National Academy of Sciences of the United States of America* 98, 5446–5451.
- Mozdzer, T.J., Zieman, J.C., 2010. Ecophysiological differences between genetic lineages facilitate the invasion of non-native *Phragmites australis* in North American Atlantic coast wetlands. *Journal of Ecology* 98, 451–458.
- Mudal, S., 1952. The chromosomes of the earthworms .1. The evolution of polyploidy. *Heredity* 6, 55–76.
- Novo, M., Almodóvar, A., Fernández, R., Giribet, G., Díaz Cosín, D.J., 2011. Understanding the biogeography of a group of earthworms in the Mediterranean basin—the phylogenetic puzzle of Hormogastridae (Clitellata: Oligochaeta). *Molecular Phylogenetics and Evolution* 61, 125–135.
- Novo, M., Almodóvar, A., Fernandez, R., Trigo, D., Diaz-Cosin, D.J., Giribet, G., 2012. Appearances can be deceptive: different diversification patterns within a group of Mediterranean earthworms (Oligochaeta, Hormogastridae). *Molecular Ecology* 21, 3776–3793.
- Nuutinen, V., Pitkanen, J., Kuusela, E., Widdom, T., Lohilahti, H., 1998. Spatial variation of an earthworm community related to soil properties and yield in a grass-clover field. *Applied Soil Ecology* 8, 85–94.
- Oksanen, J., Kindt, R., Legendre, P., O'Hara, B., Stevens, M.H.H., Oksanen, M.J., Suggests, M.A.S.S., 2007. *The Vegan Package*. Community Ecology Package.
- O'Dowd, D.J., Green, P.T., Lake, P.S., 2003. Invasional 'meltdown' on an oceanic island. *Ecology Letters* 6, 812–817.
- Pejchar, L., Mooney, H.A., 2009. Invasive species, ecosystem services and human well-being. *Trends in Ecology & Evolution* 24, 497–504.
- Perez-Losada, M., Rico, M., Marshall, J.C., Dominguez, J., 2009. Phylogenetic assessment of the earthworm *Aporrectodea caliginosa* species complex (Oligochaeta: Lumbricidae) based on mitochondrial and nuclear DNA sequences. *Molecular Phylogenetics and Evolution* 52, 293–302.
- Porco, D., Decaens, T., Deharveng, L., James, S.W., Skarzynski, D., Erseus, C., Butt, K.R., Richard, B., Hebert, P.D.N., 2013. Biological invasions in soil: DNA barcoding as a monitoring tool in a multiple taxa survey targeting European earthworms and springtails in North America. *Biological Invasions* 15, 899–910.
- R Development Core Team, 2014. *R: a Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria, ISBN 3-900051-07-0. Available at: www.R-project.org.
- Rambaut, A., Drummond, A.J., 2007. *Tracer v1.4*. Available from: <http://beast.bio.ed.ac.uk/Tracer>.
- Rauschert, E.S.J., 2006. *Competitive Interactions and Associations of the Invasive Thistles Carduus nutans and C. acanthoides*. PhD Pennsylvania State University.
- Reaser, J.K., Meyerson, L.A., Cronk, Q., De Poorter, M., Eldredge, L.G., Green, E., Kairo, M., Latala, P., Mack, R.N., Mauremootoo, J., O'Dowd, D., Orapa, W., Sastrotomo, S., Saunders, A., Shine, C., Thraillson, S., Vaiutu, L., 2007. Ecological and socioeconomic impacts of invasive alien species in island ecosystems. *Environmental Conservation* 34, 98–111.
- Revelle, W., 2011. *An overview of the psych package*. Retrieved from: <http://www.personalitytheory.org/r/book/overview.pdf>.
- Ricardo, R.P., Madeira, M., Medina, J.M.B., Marques, M.M., Furtado, A.F.S., 1977. *Esboço pedológico da ilha de S. Miguel (Açores)*. Anais do Instituto Superior de Agronomia 37, 275–385.
- Ricciardi, A., 2001. Facilitative interactions among aquatic invaders: is an "invasional meltdown" occurring in the Great Lakes? *Canadian Journal of Fisheries and Aquatic Sciences* 58, 2513–2525.
- Richard, B., Legras, M., Margerie, P., Mathieu, J., Barot, S., Caro, G., Desjardins, T., Dubs, F., Dupont, L., Decaens, T., 2012. Spatial organization of earthworm assemblages in pastures of northwestern France. *European Journal of Soil Biology* 53, 62–69.
- Ronquist, F., Huelsenbeck, J.P., 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19, 1572–1574.
- Ross, D.J., Johnson, C.R., Hewitt, C.L., Ruiz, G.M., 2004. Interaction and impacts of two introduced species on a soft-sediment marine assemblage in SE Tasmania. *Marine Biology* 144, 747–756.
- Rossi, J.P., Lavelle, P., Albrecht, A., 1997. Relationships between spatial pattern of the endogeic earthworm *Polychaeta elongata* and soil heterogeneity. *Soil Biology & Biochemistry* 29, 485–488.
- Rowell, D.L., 1994. *Soil Science: Methods and Applications*. Longman Scientific and Technical, Harlow, UK.
- Santos, R., da Conceicao, M., Pacheco, M., Dina, M., Santana, F., Muelle, H., 2005. Cyanobacteria blooms in Sete-Cidades lake - (S. Miguel Island -Azores). *Archiv fuer Hydrobiologie Supplement* 159, 393–406.
- Sharma, P.P., Giribet, G., 2012. Out of the Neotropics: Late Cretaceous colonization of Australasia by American arthropods. *Proceedings of the Royal Society B-Biological Sciences* 279, 3501–3509.
- Shekhovtsov, S.V., Golovanova, E.V., Peltek, S.E., 2014. Invasive lumbricid earthworms of Kamchatka (Oligochaeta). *Zoological Studies* 53.
- Suarez, E.R., Tierney, G.L., Fahey, T.J., Fahey, R., 2006. Exploring patterns of exotic earthworm distribution in a temperate hardwood forest in south-central New York, USA. *Landscape Ecology* 21, 297–306.
- Terhivuo, J., Saura, A., 2006. Dispersal and clonal diversity of North-European parthenogenetic earthworms. *Biological Invasions* 8, 1205–1218.
- Thompson, J.D., Gibson, T.J., Plewniak, F., Jeanmougin, F., Higgins, D.G., 1997. The CLUSTAL X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Research* 25, 4876–4882.
- Uvarov, A.V., 2009. Inter- and intraspecific interactions in lumbricid earthworms: their role for earthworm performance and ecosystem functioning. *Pedobiologia* 53, 1–27.
- Vandel, A., 1928. La parthénogénèse géographique: contribution à l'étude biologique et cytologique de la parthénogénèse naturelle. *Bulletin Biologique de la France et de la Belgique* 62, 164–281.

- VanDyke, K.A., Lockwood, J.A., Kazmer, D.J., 2004. Association of genetic lineages with ecological features in a polyphagous montane grasshopper species. *Journal of Orthoptera Research* 13, 205–209.
- Viktorov, A.G., 1997. Diversity of polyploid races in the family Lumbricidae. *Soil Biology & Biochemistry* 29, 217–221.
- Vila, M., Basnou, C., Pysek, P., Josefsson, M., Genovesi, P., Gollasch, S., Nentwig, W., Olenin, S., Roques, A., Roy, D., Hulme, P.E., Andriopoulos, P., Arianoutsou, M., Augustin, S., Bacher, S., Bazos, I., Bretagnolle, F., Chiron, F., Clergeau, P., Cochard, P.O., Cocquempot, C., Coeur d'Acier, A., David, M., Delipetrou, P., Desprez-Loustau, M.L., Didziulis, V., Dorkeld, F., Essl, F., Galil, B.S., Gasquez, J., Georgioudis, K., Hejda, M., Jarosik, V., Kark, S., Kokkoris, I., Kuhn, I., Lambdon, P.W., Lopez-Vaamonde, C., Marcer, A., Migeon, A., McLoughlin, M., Minchin, D., Navajas, M., Panov, V.E., Pascal, M., Pergl, J., Perglova, I., Pino, J., Poblisaj, K., Rabitsch, W., Rasplus, J.Y., Sauvard, D., Scalera, R., Sedlacek, O., Shirley, S., Winter, M., Yannitsaros, A., Yart, A., Zagatti, P., Zikos, A., Partners, D., 2010. How well do we understand the impacts of alien species on ecosystem services? A pan-European, cross-taxa assessment. *Frontiers in Ecology and the Environment* 8, 135–144.
- Viveiros, F., Ferreira, T., Silva, C., Gaspar, J.L., 2009. Meteorological factors controlling soil gases and indoor CO₂ concentration: a permanent risk in degassing areas. *Science of the Total Environment* 407, 1362–1372.
- Walsh, C., Mac Nally, R., 2011. Package 'hier.part'. <http://cran.r-project.org/web/packages/hier.part/hier.part.pdf>.
- Woolfit, M., Bromham, L., 2005. Population size and molecular evolution on islands. *Proceedings of the Royal Society B-Biological Sciences* 272, 2277–2282.